# Aurora Australis Marine Science Cruise AU1603 - Oceanographic Field Measurements and Analysis

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#### **1 INTRODUCTION**

Oceanographic measurements were collected aboard Aurora Australis cruise au1603, voyage 3 2015/2016, from 11th January to ~24th February 2016. The cruise commenced with the K-AXIS project, the major marine science component of the cruise. This was the Australian component (P.I.'s Andrew Constable, Steve Rintoul and others) of a combined biological and oceanographic study in the vicinity of the Kerguelen Axis. After conclusion of marine science work the ship went to Mawson for a resupply. During a storm on 24th February the ship broke free of its mooring lines and ran aground on the rocks at West Arm in Horseshoe Harbour, thus ending the cruise. Expeditioners were eventually taken to Casey on the Shirase, then flown home. Meanwhile the Aurora Australis was refloated and sailed to Fremantle, then on to Singapore for repairs.

This report discusses the oceanographic data from CTD operations on the cruise. A total of 47 CTD vertical profile stations were taken on the cruise (Table 1). Over 850 Niskin bottle water samples were collected for the measurement of salinity, dissolved oxygen, nutrients (phosphate, nitrate+nitrite and silicate), dissolved inorganic carbon (i.e.  $TCO<sub>2</sub>$ ), alkalinity, POC and PN, and biological parameters, using a 24 bottle rosette sampler. A UVP particle counter/camera system was attached to the CTD package (P.I. Emmanuel Laurenceau). A separate trace metal rosette system was deployed from the trawl deck (P.I. Andrew Bowie). Upper water column current profile data were collected by a ship mounted ADCP, and meteorological and water property data were collected by the array of ship's underway sensors. Eight drifting floats were deployed over the course of the cruise.

Processing/calibration and data quality for the main CTD data are described in this report. Underway sea surface temperature and salinity data are compared to near surface CTD data. CTD station positions are shown in Figure 1, while CTD station information is summarised in Table 1. Float deployments (5 x Argo/Apex, 2 x SOCCOM and 1 x Provor) are summarised in Table 10. Further cruise itinerary/summary details can be found in the voyage leader report (Australian Antarctic Division unpublished report: Voyage 3 2015-2016, RSV Aurora Australis, Voyage Leader's report).

#### **2 CTD INSTRUMENTATION**

SeaBird SBE9plus CTD serial 704, with dual temperature and conductivity sensors and a single SBE43 dissolved oxygen sensor (serial 0178, on the primary sensor pump line), was used, mounted on a SeaBird 24 bottle rosette frame, together with a SBE32 24 position pylon and 24 x 10 litre General Oceanics Niskin bottles. The following additional sensors/instruments were mounted:

\* Wetlabs ECO-AFL/FL fluorometer serial 756 (analog range 2)

- \* Biospherical Instruments PAR sensor QCP2300HP, serial 70110
- \* Wetlabs C-star transmissometer serial 899DR
- \* Tritech 200 kHz altimeter serial 237622
- \* Tritech 500 kHz altimeter serial 76031
- \* UVP/camera system and lighting (a few stations only)

CTD data were transmitted up a 8 mm seacable to a SBE11plusV2 deck unit, at a rate of 24 Hz, and data were logged simultaneously on 2 PC's using SeaBird data acquisition software "Seasave" (version unknown).

The CTD deployment method was as follows:

- \* CTD initially deployed down to ~10 to 20 m
- \* after confirmation of pump operation, CTD returned up to just below the surface (depth dependent on sea state)
- \* after returning to just below the surface, downcast proper commenced

Pre cruise temperature, conductivity and pressure calibrations were performed by SeaBird (Table 2) (July 2015). The SeaBird calibration for the SBE43 oxygen sensor was used for initial data display only. Manufacturer supplied calibrations were used for the fluorometer, transmissometer, PAR and altimeter. Final conductivity and dissolved oxygen calibrations derived from in situ Niskin bottle samples are listed later in the report. Final transmissometer data are referenced to a clean water value. UVP data are not discussed.

#### **3 PROBLEMS ENCOUNTERED**

Running aground of the ship at Mawson was obviously the most serious incident on the cruise. On the equipment side, CTD operations went relatively smoothly, with few significant equipment problems. Notable problems (on a lesser scale of drama than grounding of the ship) were as follows:

\* Grease contamination of the rosette was a problem over several days (first noted on station 23). The grease, from a winch on the trawldeck, was inadvertently being brought into the CTD room and daubed over the Niskins and rosette frame. Grease was removed with paper towel prior to station 24. After station 24 the Niskins and frame were thoroughly cleaned with isopropyl alcohol, and during station 25 (while CTD was in the water) the CTD room was thoroughly cleaned with hot water.

\* There was difficulty interpreting altimeter readings for station 1, and bottom of cast was at ~79 m above the bottom.

\* At station 12, the CTD was initially deployed with the fluorometer cap left on. The package was recovered then redeployed with the cap removed.

\* Niskin 5 leaked for many stations, and was eventually replaced prior to station 17.

\* Station 18 was moved by ~3miles due to heavy ice. Station 20 was also moved, due to ice.

\* Station 30: some delay due to problems with gantry. Grease cleaned from rosette and CTD room deck while gantry was being repaired.

\* Station 33: the trolley broke after retrieval, so the CTD couldn't be shifted inboard. Sampling was done with the CTD hard up against the CTD door.

\* For station 35 onwards, in many cases the CTD was taken to near bottom but samples were only collected in the upper water column. It will therefore not be possible to reliably calibrate much of the CTD oxygen data for these stations, in particular for stations 38-40 and 42-43.

\* Station 44: pump took a long time to come on, and reboots were required (PC's and deck unit). As a result the CTD was already in the water by the time logging was successfully started.

\* Station 47: when CTD at 55 dbar on upcast, returned back down to 200 dbar to fire a bottle.

### **4 CTD DATA PROCESSING AND CALIBRATION**

Final processing of the CTD data was done in Hobart (only limited processing was possible at sea). The first processing step is application of a suite of the SeaBird "Seasoft" processing programs to the raw data, in order to:

\* convert raw data signals to engineering units

\* remove the surface pressure offset for each station

\* realign the oxygen sensor with respect to time (note that conductivity sensor alignment is done by the deck unit at the time of data logging)

\* remove conductivity cell thermal mass effects

\* apply a low pass filter to the pressure data

\* flag pressure reversals

\* search for bad data (e.g. due to sensor fouling etc)

Further processing and data calibration were done in a UNIX environment, using a suite of fortran and matlab programs. Processing steps here include:

\* forming upcast burst CTD data for calibration against bottle data, where each upcast burst is the average of 10 seconds of data centered on each Niskin bottle firing

\* merging bottle and CTD data, and deriving CTD conductivity calibration coefficients by comparing upcast CTD burst average conductivity data with calculated equivalent bottle sample conductivities \* forming pressure monotonically increasing data, and from there calculating 2 dbar averaged downcast CTD data

\* calculating calibrated 2 dbar averaged salinity from the 2 dbar pressure, temperature and conductivity values

\* deriving CTD dissolved oxygen calibration coefficients by comparing bottle sample dissolved oxygen values (collected on the upcast) with CTD dissolved oxygen values from the equivalent 2 dbar downcast pressures

Full details of the data calibration and processing methods are given in Rosenberg et al. (unpublished) referred to hereafter as the  $CTD$  methodology. Additional processing steps are discussed below in the results section. For calibration of the CTD oxygen data, whole profile fits were used for shallower stations, while split profile fits were used for deeper stations.

Final station header information, including station positions at the start, bottom and end of each CTD cast, were obtained from underway data for the cruise (see section 6 below). Note the following for the station header information:

\* All times are UTC.

\* "Start of cast" information is at the commencement of the downcast proper, as described above.

\* "Bottom of cast" information is at the maximum pressure value.

\* "End of cast" information is when the CTD leaves the water at the end of the cast, as indicated by a drop in salinity values.

\* All bottom depth values are corrected for local sound speed, where sound speed values are calculated from the CTD data at each station.

\* "Bottom of cast" depths are calculated from CTD maximum pressure (converted to depth) and altimeter values at the bottom of the casts.

Lastly, data were converted to MATLAB format, and final data quality checking was done within MATLAB.

#### **5 CTD AND BOTTLE DATA RESULTS AND DATA QUALITY**

Data from the secondary CTD sensor pair (temperature and conductivity) were used for the whole cruise. Suspect CTD 2 dbar averages are listed in Table 8. Data from the test cast (station 0) were not processed.

#### **5.1 Conductivity/salinity**

The conductivity calibration and equivalent salinity results for the cruise are plotted in Figures 2 and 3, and the derived conductivity calibration coefficients are listed in Tables 3 and 4. Station groupings used for the calibration are included in Table 3. International standard seawater batch number P158 (expiry 25th March 2018) was used for salinometer standardisations.

Guildline Autosal serial 62549 was used for the whole cruise, with analyses taking place in the skylab. Salinometer performance was stable, with lab temperature averaging 20.5  $\pm$ 0.8 °C during analyses.

Overall salinity accuracy for the cruise is within 0.002 (PSS78). Note that very low near surface salinities occurred for some stations (most notably station 2 to 4, 12, and 17 to 22). These features are typically accompanied by steep vertical gradients (often with little, if any, surface mixed layer), and for such cases it's difficult to obtain salinity samples suitable for calibration of the CTD data. As a result, very few low salinity sample values were included in the calibrations. The manufacturer claims little significant non-linearity in the conductivity cell response, so any additional inaccuracy for the lowest salinity values, i.e. salinity < 33.2 (PSS78), is assumed to be small (of the order 0.001 at most over the implied calibration extrapolation range).

Pressure dependent salinity residuals are evident for most cruises, due to pressure effects on the glass conductivity cells (SeaBird tech, personal communication). For this cruise the residuals were small, and where they occurred they were of the order 0.002 (PSS78) or less over the whole vertical profile.

Close inspection of the vertical profiles of the bottle-CTD salinity difference values reveals a slight biasing for a few stations, mostly of the order 0.001 (PSS78), as follows:



This is most likely due to a combination of factors, including salinometer performance. There is no significant diminishing of overall CTD salinity accuracy from this apparent biasing.

Bad salinity bottle samples (not deleted from the data files) are listed in Table 9.

\* Salinity samples for station 44 to 47 were unreliable, and conductivity calibrations for these stations were extrapolated from previous stations. For station 44, samples do not compare well with CTD data. For stations 45 to 47, samples do not seem to correspond with bottle firings (there may have been a mixing up of crates, or some error in data entry).

\* For stations 46 and 47, secondary conductivity data were suspect on the upcast over the following intervals: 315 to 91 dbar (station 46), and 268 to 70 dbar (station 47). There may have been something on the frame interfering with flow past the sensors, but this is inconclusive.

#### **5.2 Temperature**

Temperature differences between the primary and secondary CTD temperature sensors ( $T_p$  and  $T_s$ ) respectively), from data at Niskin bottle stops, are shown in Figure 4. The difference  $T_s - T_p$ , at an average of  $0.0005^{\circ}$ C over the whole profile (Figure 4a), is within the manufacturer quoted sensor  $accuracy of  $0.001^{\circ}C$ .$ 

#### **5.3 Pressure**

Surface pressure offsets for each cast (Table 5) were obtained from inspection of the data before the package entered the water. Pressure spiking, a problem on some previous cruises, did not occur. For station 44, the CTD was already in the water by the time logging was successfully started, so the surface pressure offset value from station 45 was used.

#### **5.4 Dissolved oxygen**

CTD oxygen data were calibrated as per the CTD methodology, with profiles deeper than 1400 dbar calibrated as split profile fits, and profiles shallower than 1400 dbar (i.e. stations 7 to 9, 21, 24, 26, 35- 37, 40 and 45-47) calibrated as whole profile fits. Additional stations also calibrated as whole profile fits were: stations 31 and 41 (with a maximum pressures of 1561 and 1738 dbar respectively); and stations 38, 39, 42 and 43 (no bottle samples for calibration of deep part of profile).

Calibration results are plotted in Figure 5, and the derived calibration coefficients are listed in Table 6. Overall the calibrated CTD oxygen agrees with the bottle data to within 1% of full scale (where full scale is ~400 µmol/l above 1500 dbar, and ~250 µmol/l below 1500 dbar).

\* For stations 1 to 5, there's a small error in the temperature of the iodate reagent used to calculate the amount of reagent dispensed. Any resulting error in the bottle analyses is assumed to be small (less than 1% full scale).

\* For station 13, reagent 2 was added using a hand pipette.

\* For station 26 oxygen sampling, reagents were accidentally dispensed in reverse order (i.e. reagent 2 followed by reagent 1). Samples were not analysed (therefore no CTD oxygen data).

\* Stations 30-34: CTD oxygen spikes around ~200 dbar on the downcast each time.

\* For station 35 onwards, in many cases the CTD was taken to near bottom but samples were only collected in the upper water column. It was therefore not be possible to reliably calibrate much of the CTD oxygen data for these stations, in particular for the full depth stations 38-40 and 42-43.

\* For station 47, CTD oxygen data for 2 to 50 dbar are suspect.

#### **5.5 Fluorescence, PAR, transmittance, altimeter**

All fluorescence, PAR and transmittance data have a manufacturer supplied calibration (Table 2) applied to the data, with transmittance values referenced to clean water. In the CTD 2dbar averaged data files, both downcast and upcast data are supplied for these sensors; and the data are strictly 2 dbar averages (as distinct from other calculations used in previous cruises i.e. au0703, au0803 and au0806).

The PAR calibration coefficients in Table 2 were calculated from the manufacturer supplied calibration sheet, using the method described in the following SeaBird documents: page 53 of SeaSave Version 7.2 manual; Application Note No. 11 General; and Application Note No. 11 QSP-L. The PAR calibration "offset" value (Table 2) was derived from deep water voltage values from the trials cruise in December 2015.

Maximum transmittance values are slightly less than the expected 100%, and the station means drift downward over the cruise by ~4%. The former is most likely a small calibration error, while the later is most likely due to a lack of cleaning of the sensor windows. For stations 1 and 2, values are slightly higher than 100% - most likely a small calibration error again. Station 28 downcast transmittance data were very low (and these data have been removed), most likely due to contamination of sensor windows.

The usual altimeter "artefacts" (as seen on previous cruises) were observed on occasion, with false bottom readings often observed before coming within nominal altimeter range. For station 1, minimum altimeter height above the bottom was ~79 m. due to uncertainty in interpretation of altimeter readings.

#### **5.6 Nutrients**

All nutrients were frozen and returned to Hobart, with analysis taking place at CSIRO between November 2016 and March 2017. As a result of this long delay the data were approached with great caution. Data from the Roger Revelle 2016 cruise along CLIVAR I8S transect (expocode 33RR20160208) (Figure 8) were extremely valuable for verification and quality control of the K-AXIS data. Note that K-AXIS nutrients were first converted to gravimetric units (umol/kg) to allow direct comparison to the Revelle data. Laboratory temperature used for this conversion was 20°C. Overall, K-AXIS silicate and nitrate+nitrite data comparisons to the Revelle data are reasonable (Figures 10 and 9 respectively). K-AXIS phosphates however are noisier, and are consistently offset from the Revelle data by ~0.05 µmol/kg (Figure 7). From profile comparisons and inspection of nitrate+nitrite versus phosphate data (Figure 6), a significant number of phosphate values were initially flagged as either suspect or bad. Taking into account the offset from the Revelle data, all K-AXIS phosphates are considered suspect, and have been flagged accordingly. Note that the bottle data files include nutrient flags, with the following flag values:

- $1 =$  below detection threshold (applies to some ammonia & nitrite data from station 3 and 44)
- $2 = a$ ood
- $3 =$ suspect
- $4 = bad$
- $9 = no$  data

Quality control decisions were sometimes difficult, in particular for isolated southern stations following a frontal feature (e.g. stations 21 and 22). In general, values are only flagged as bad (flag value 4) if they are out by at least ~10% of full scale (where full scale  $\approx 3.0$ , 40 and 150 umol/l respectively for phosphate, nitrate+nitrite and silicate), or if they are obvious outliers. However for this data set, many nitrate and phosphate data points that would usually be flagged as bad were left with a suspect flag only – again, due to the difficulty of QC decisions for this cruise. Data flagged as suspect were values deemed to be out by at least ~3% (in most cases) of full scale. In some cases (e.g. phosphate and nitrate for station 21 bottles 1, 4-6 and 10, and station 22 bottles 7 and 9) it is unknown whether apparent spikes in the profile are real features or not – flag values were left as 2 for these.

Note that nutrient samples assigned a flag value of 3 or 4 are listed in the series of text files bad\* (e.g. badphos) and suss\* (e.g. sussphos) included with the data set. The phosphate samples listed in the file sussphos are judged as outliers within the vertical profiles – this information may be of use, and is additional to the overall suspect label for all the phosphate data.

For stations 3 and 44, nutrients were analysed by hydrochemistry groups from both CSIRO and SCRIPPS (results not discussed here). The CSIRO silicate data for these 2 stations are all bad, so the SCRIPPS data (converted back to µmol/l units from the original µmol/kg, using an assumed lab temperature of  $20^{\circ}$ C) have been used instead.

#### **6 UNDERWAY MEASUREMENTS**

Underway data were logged to an Oracle database on the ship. Quality control for the cruise was largely automated. 12 kHz bathymetry data were not quality controlled, and these data in the underway file are from the sounder's automatic bottom detection – as a result, the data are scattered and often incorrect. Bottom depths at CTD stations were carefully extracted (i.e. good values only were used), and corrected for the local sound speed at each station.

10 second instantaneous underway data are contained in the file au1603.ora as column formatted text; and in the file au1603ora.mat as matlab format. Data from the hull mounted underway temperature sensor  $(T_{\text{dis}})$  and the underway thermosalinograph salinity ( $S_{\text{dis}}$ ) are compared to CTD temperature and salinity data at 8 dbar (Figures 11 and 12). Offset corrections are sufficient in both cases (Figure 12). Note that for salinity, underway salinity values are too low for decimal days 30 to 38 (Figure 11) i.e. CTD stations 18 to 31, possibly due to fouling of the sensor or the flow inlet. These data have been excluded from the offset calculation. Also note that the offset corrections have not been applied to the underway data files.

#### **7 FILE FORMATS**

Data are supplied as column formatted text files, or as matlab files, with all details fully described in the README file included with the data set. Note that all dissolved oxygen and nutrient data in these file versions are in units of umol/l.

The data are also available in WOCE "Exchange" format files. In these file versions, dissolved oxygen and nutrient data are in units of umol/kg. For density calculation in the volumetric to gravimetric units conversion, the following were used:

dissolved oxygen – in situ temperature and CTD salinity at which each Niskin bottle was fired; zero pressure

nutrients – laboratory temperature ( $20^{\circ}$ C), and in situ CTD salinity at which each Niskin bottle was fired; zero pressure

#### **REFERENCES**

Rosenberg, M., Fukamachi, Y., Rintoul, S., Church, J., Curran, C., Helmond, I., Miller, K., McLaughlan, D., Berry, K., Johnston, N. and Richman, J., unpublished. Kerguelen Deep Western Boundary Current Experiment and CLIVAR I9 transect, marine science cruises AU0304 and AU0403 - oceanographic field measurements and analysis. ACE Cooperative Research Centre, unpublished report. 78 pp.

#### **ACKNOWLEDGEMENTS**

Thanks to all scientific personnel who participated in the cruise, and to the crew of the RSV Aurora Australis. Special thanks to the oceanography team for a great job collecting the data.

**Table 1: Summary of station information for cruise au1603. All times are UTC; "alt" = minimum altimeter value (m), "maxp" = maximum pressure (dbar).** 





**Table 2: CTD calibration coefficients and calibration dates for cruise au1603. Note that platinum temperature calibrations are for the ITS-90 scale. Pressure slope/offset, temperature, conductivity and oxygen values are from SeaBird calibrations. Fluorometer and PAR values are manufacturer supplied (with the PAR offset value updated from dark voltage values observed on the trials cruise in December 2015). Transmissometer values are a rescaling of the manufacturer supplied coefficients to give transmittance as a %, referenced to clean water. For oxygen, the final calibration uses in situ bottle measurements (the manufacturer supplied coefficients are not used).** 



**Table 3: CTD conductivity calibration coefficients for cruise au1603. F1 , F2 and F3 are respectively conductivity bias, slope and station-dependent correction calibration terms. n is the number of samples retained for calibration in each station grouping;** σ **is the standard deviation of the conductivity residual for the n samples in the station grouping.** 



**Table 4: Station-dependent-corrected conductivity slope term (F2 + F3 . N), for station number N, and F2 and F3 the conductivity slope and station-dependent correction calibration terms respectively, for cruise au1603.**



**Table 5: Surface pressure offsets (i.e. poff, in dbar) for cruise au1603. For each station, these values are subtracted from the pressure calibration "offset" value in Table 2. Note: for station 44, pressure offset from station 45 used.**



**Table 6: CTD dissolved oxygen calibration coefficients for cruise au1603: slope, bias, tcor ( = temperature correction term), and pcor ( = pressure correction term). dox is equal to 2.8**σ **, for** σ **as defined in the CTD Methodology. For deep stations, coefficients are given for both the shallow and deep part of the profile, according to the profile split used for calibration (see section 5.4 in the text); whole profile fit used for stations shallower than 1400 dbar (i.e. stations with only "shallow" set of coefficients in the table), plus stations 31, 38-39 and 41-43.** 



**Table 7: Missing data points in 2 dbar-averaged files for cruise au1203. "x" indicates missing data for the indicated parameters: T=temperature; S/C=salinity and conductivity; O=oxygen; F=fluorescence downcast; PAR=photosynthetically active radiation downcast;** 

**TR=transmittance downcast; F\_up=fluorescence upcast; PAR\_up=photosynthetically active radiation upcast; TR\_up=transmittance upcast.** 

**Note: 2 and 4 dbar values not included here - 2 dbar value missing for most casts, 4 dbar value missing for many casts.** 



#### **Table 8: Suspect CTD 2 dbar averages (not deleted from the CTD 2 dbar average files) for the indicated parameters, for cruise au1603.**





# **Table 10: Summary of APEX Argo, SOCCOM and Provor float deployments on cruise au1603.**





**Figure 1: CTD station positions and ship's track for cruise au1603.** 



**Figure 2:** Conductivity ratio  $c_{\text{btl}}/c_{\text{cal}}$  versus station number for cruise au1603. The solid line **follows the mean of the residuals for each station; the broken lines are** ± **the standard**  deviation of the residuals for each station.  $c_{cal}$  = calibrated CTD conductivity from the CTD upcast burst data; c<sub>btl</sub> = 'in situ' Niskin bottle conductivity, found by using CTD pressure and **temperature from the CTD upcast burst data in the conversion of Niskin bottle salinity to conductivity.** 



Figure 3: Salinity residual (S<sub>btl</sub> - S<sub>cal</sub>) versus station number for cruise au1603. The solid line is **the mean of all the residuals; the broken lines are** ± **the standard deviation of all the residuals. s**<sub>cal</sub> = calibrated CTD salinity;  $s_{\text{btl}}$  = Niskin bottle salinity value.



**Figure 4: Difference between secondary and primary temperature sensors with (a) pressure, and (b) temperature. Data are from the upcast CTD data bursts at Niskin bottle stops.** 



**Figure 5: Dissolved oxygen residual (obtl - ocal) versus station number for cruise au1603. The solid line follows the mean residual for each station; the broken lines are** ± **the standard**  deviation of the residuals for each station. o<sub>cal</sub>=calibrated downcast CTD dissolved oxygen; **o**<sub>btl</sub> = Niskin bottle dissolved oxygen value. Note: values outside vertical axes are plotted on **axes limits.** 



**Figure 6: CTD Station positions for au1603 (K-AXIS) and Roger Revelle I8s 2016 cruise.** 



**Figure 7: Nitrate+nitrite versus phosphate data, for au1603 (K-AXIS, whole cruise) and Roger Revelle I8S 2016 (stations 7 to 28). Note that K-AXIS data in the plot have been converted to units µmol/kg. Also note that K-AXIS phosphate shown as suspect are prior to flagging the entire phosphate data set as suspect.**



**Figure 8: Phosphate profiles for au1603 (K-AXIS, whole cruise), and Roger Revelle I8S 2016 (stations 7 to 28). Note that K-AXIS phosphate shown as suspect are prior to flagging the entire phosphate data set as suspect.**



**Figure 9: Nitrate+nitrite profiles for au1603 (K-AXIS, whole cruise), and Roger Revelle I8S 2016 (stations 7 to 28). Note that K-AXIS data in the plot have been converted to units µmol/kg.**



**Figure 10: Silicate profiles for au1603 (K-AXIS, whole cruise), and Roger Revelle I8S 2016 (stations 7 to 28). Note that K-AXIS data in the plot have been converted to units µmol/kg.**



**Figure 11: au1603 comparison of underway temperature and salinity data to CTD data, with time.** 



**Figure 12a and b: au1603 comparison between (a) CTD and underway temperature data (i.e. hull mounted temperature sensor), and (b) CTD and underway salinity data. Note: dls refers to underway data. Note that due to the large scatter these corrections have not been applied to the underway data.** 

# **APPENDIX 1 KAXIS HYDROCHEMISTRY REPORT**

(at the time of writing: this appendix is for salinity analyses only)

#### Personnel

Analysis-Ruth Eriksen Sampling- Katherine Tattersall, Delphi Ward, Stuart Corney, Christine Weldrick, plus volunteers Justin Phebey and Tom Clarke

Key questions/outputs (from voyage plan)

- Calibration of dissolved oxygen and salinity sensors for 47 CTD casts through on-board analysis of samples drawn from Niskins (Rintoul/Rosenberg)
- Collection and preservation of nutrient samples for analysis in Hobart (Rintoul/Rosenberg)
- Measurement of dissolved oxygen concentrations in  $CO<sub>2</sub>$  manipulation experiments (King/Kawaguchi)

#### Strategy behind sampling design

• Sampling was conducted in accordance with standard practice for chemical oceanography. Depths for sampling were based on examination of the down-cast by the CTD watch, with input from groups requiring water as per the water budget. Detailed notes provided by Mark Rosenberg and Steve Rintoul guided decision making about cast deployments and subsequent water masses sampled.

#### Methods

**Salinity** 

- Salinity samples were collected according to the CSIRO Procedure S2 V01, using new "square" clear glass bottles that had been conditioned with surface seawater collected on earlier voyages onboard Investigator.
- Samples were analysed using a Guildline Model 8400B "Autosal" salinometer (Units 62548 and 62549)
- Analysis protocols were drawn from CSIRO procedures (G3 V01), and the Guildline manual
- Additional information on analysis was drawn from Kawano GO-SHIP (IOCCP Report No 14, 2010) and the original CSIRO Hydrochemistry Manual (Cowley et al 1999)
- Instrument standardisation was undertaken using IAPSO Standard Seawater, Batch P158 (Expiry 25.3.2018)
- Laboratory temperature was set to 20  $^{\circ}$ C by the ships engineer, as per hydrochem report by Craig Neill on the previous year's Totten Glacier voyage.
- Laboratory temperature was logged using a HOBO data logger, at 5 min intervals for the duration of the voyage (Figure 1). Lab temperature averaged 20.5  $\pm$ 0.8 °C during the marine science period of the voyage. Temperature dropped immediately after the grounding to 17.8  $^{\circ}$ C.
- Water bath temperature in the salinometers was logged using a high precision temperature probe constructed by CSIRO for the voyage (Figure 2). Data was logged at 2 min intervals using the HYDRO1 software developed by CSIRO. The temperature probe was calibrated prior to the voyage by the O&A Oceanographic Calibration Facility on 21/12/2015 over the range -1.458 to 32.042 °C.
- Duplicates were not routinely analysed, on advice from Mark Rosenberg. An initial set of duplicates was collected on CTD 000 and 001 for the purposes of training the CTD watch in sampling technique.



**Figure 1: Laboratory temperature log, Skylab V3 from January 13 – February 29 2016** 



**Figure 2: 24 hr average temperature± 0.001 <sup>o</sup>C (dotted lines) for Guildline salinometer 62549, KAXIS -V3. Bars represent 24 hour range. Temperature logger in Guildline 62548 23rd January to 27th January. Logger removed 2nd Feb to calibrate DO sensors.** 

#### Sample processing

- All samples were processed on-board, as the salinometer appeared stable and reproducible, compared to previous recent voyages (see report by Craig Neill).
- Samples were left to equilibrate for 24 hours in the Skylab before analysis.
- At the start of each run, the instrument was first flushed with surface seawater stored in a 20L carboy, until a stable reading was achieved. Next the flow-cell was flushed with open IAPSO seawater (i.e the previous days IAPSO bottle/s).
- The instrument was then calibrated using 1 or 2 (occasionally 3) new bottles of IAPSO Standard (batch P158) until 3 repeat reads were within acceptable limits. Water bath temp, instrument drift, zero and standby values were checked repeatedly during each run.
- The cell was flushed at least 3 times with a new sample before taking duplicate reads.
- Data was manually processed using the Excel spreadsheet "Saltsheet.xlsm" provided by the Hydrochemistry group, as no Guildline data logger was available for the voyage.
- Preliminary data analysis onboard included plotting lab salinity against scan salinity during the upcast, as a first pass to check for outliers, typos, or sampling mix-ups.
- Data was then passed to Stuart Corney for first stage processing according to Mark Rosenbergs protocols.

Sample summary (how many samples for each method, location, time)

- A sample summary table for all hydrochemistry samples (salinity, dissolved oxygen and nutrients) is presented as Table 1. Note that nutrient samples were collected in triplicate.
- Salinity samples were also collected from the TMR, however the majority of these were analysed in Hobart due to the grounding (see report by C. Schallenberg for more details).
- A summary of analysis and preliminary data checks conducted on –board is included in

Lab set-up (instrumentation, bench configuration, filtration racks, storage)\

Salinity analysis was set up on the central bench in the Skylab, with one salinometer at either end. This allowed standards, samples and the data logging gear to be positioned centrally and accessed from either instrument.

Sample bottles (outgoing) were stacked by the Skylab door, and sample bottles (incoming) were stacked by the salinometers, rotating up onto the benches for 24 hours prior to analysis.

A sample entry/exit log was maintained so that the CTD watch could check crates of bottles in and out. This was useful for analysis, as it allowed easy identification of how long samples had been equilibrating in the Sky lab.

Temperature loggers were placed beside the salinometer (Figure 1), in the water bath ( Figure **2**) and near the DO system.

#### Recommendations (what would you do differently)

- Sufficient salinity sample bottles were taken on V3 so that it would be possible to bring all samples back to Hobart for analysis in the event of instrument failure. The sample bottles supplied for this purposed were the type used to supply the IAPSO Standard seawater, as they have crimpable lids which should provide a robust seal to deal with delayed analysis.
- This type of bottle has a narrow neck, and is time-consuming to sample (draining is slow through the small neck) compared to standard bottles.
- All the bottles (~1000) in total were pre-conditioned with surface seawater prior to the voyage.
- There was some questions from Customs officials during the quarantine process conducted in Freemantle as to the necessity to declare these "samples". I had confirmed with the DVL earlier in the voyage that the bottles were filled with coastal water (Maria Is water collected by the Hydrochemistry group during routine IMOS sampling), and it was decided that it was not necessary to include these in the Quarantine declaration. It would be useful to confirm this before the next voyage, and label the boxes accordingly.



#### **Table 1: Summary of analytical conditions and preliminary data checks for salinity analyses.**  *Note all calibrations using OSIL batch P158 (K15 =0.99970)*

 $<sup>1</sup>$  Logger installed on 62548.</sup>

<sup>2</sup> Logger removed to trouble shoot DO rig<br><sup>3</sup> Plus bottom for genetics<br><sup>4</sup> Calibration good and no drift observed for Stations 44- 47, but overall decrease in level of agreement between CTD scan data and bottle data. Analysed post- grounding, on a slight angle

# **Table 2: Summary of equipment and consumables used for MS Trials and V3-KAXIS**



- The results of storage trials conducted by the Hydrochemistry group suggested that the results obtained from the "IAPSO" style bottles were slightly lower than results obtained from the regular salinity bottles, so there was some concern that bringing (some) samples back to Hobart for analysis might result in an offset in the data set. The offset is presumably due to the IAPSO bottles providing a superior seal, but this should be investigated further/confirmed. There was no capacity for me to pursue sample bottle comparisons during V3, although we did trial duplicate sampling so that the CTD watch were familiar and comfortable with the crimping process.
- For this reason, all samples were analysed on-board, with the caveat that analysis would stop if instrument performance deteriorated. This resulted in an intense workload for the sole hydrochemist at times.
- The instrument calibration appeared good after the grounding, so the remaining samples (CTDs 45-47) were analysed post-grounding. There was a great deal of uncertainty during those few days about the likelihood of the samples returning to Hobart, and the time that may take, so I elected to analyse all outstanding samples. It should be noted that the lab temp dropped, but that the salinonometer bath temp was stable.
- Some electrical noise from winches during trawls was observed on both trials and V3. Work flow was organised so that generally, Dos were analysed during trawling operations, and Salinities were analysed whilst we were on station doing CTD casts, or transiting

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Bottle Depth Feature Oxygen Salinity Nutrients Depth Feature O <sup>24</sup> <sup>5</sup> Surface <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>5</sup> Surface <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>5</sup> Surface <sup>5</sup> Surface <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>5</sup> Surface <sup>1</sup> <sup>1</sup> <sup>1</sup> $\overline{1}$ 23 5 Surface 1 1 1 1 5 Surface 1 5 Surface 1 1 1 1 5 Surface 1 1 1 1 1 1 1 5 Surface 1 1 1 1  $\overline{1}$ 22 15 DCM 1 1 1 10 DCM 1 1 1 1 15 DCM <sup>15</sup> DCM <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>15</sup> DCM <sup>1</sup> <sup>1</sup> <sup>1</sup> 21 15 DCM 1 1 1 1 10 DCM <sup>15</sup> DCM <sup>15</sup> DCM <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>15</sup> DCM <sup>1</sup> <sup>1</sup> <sup>1</sup> 20 15 DCM<br>19 20 MLD M 1 20 1 1 1 1 5 DCM 1 1 1 1 1 20 NM 19 20 MLD 1 1 1 1 30 1 1 1 20 MLD 1 1 1 20 MLD 1 1 1 20 MLD 1 1 25  $\overline{1}$ 18 30 1 1 1 1 35 1 1 1 30 1 1 1 35 1 1 1 1 35 MLD 1 1 1  $\mathbf{1}$ 17 40 1 1 1 1 40 MLD 1 1 1 40 1 1 1 1 1 50 Tmin 1 1 1 45 1 1 1 1  $\mathbf{1}$ <sup>16</sup> <sup>50</sup> CHL0 <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>75</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>60</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>75</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>60</sup> Tmin <sup>1</sup> <sup>1</sup> <sup>1</sup> $\overline{1}$ <sup>15</sup> <sup>75</sup> Tmin <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>75</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>80</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\overline{1}$ <sup>14</sup> <sup>100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>150</sup> O2, Tmax <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>150</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>125</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\overline{1}$ 13 150 1 1 1 1 200 1 1 1  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<sup>1500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>1000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>1500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>1250</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\overline{1}$ <sup>6</sup> <sup>1250</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>1750</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>1500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>1800</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>1500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\overline{1}$ <sup>5</sup> <sup>1500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\,$  1  $\,$ <sup>4</sup> <sup>1750</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\mathbf 1$ <sup>3</sup> <sup>2100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2700</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>3000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>3000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>3000</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\mathbf{1}$ 2 2480 1 1 1 1 2830 1 1 1 3200 1 1 1 3115 1 1 1 3270 1 1 1 1  $\mathbf{1}$ <sup>1</sup> <sup>2530</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>2885</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>3260</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>3165</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>3320</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup>Bottle Depth Feature Oxygen Salinity Nutrients Depth Feature Oxygen Salinity Neture Oxygen Salinity Nutrients Depth Geature Oxygen Salinity Nutrients Depth Feature Oxygen Salinity Nutrients 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DCM/MLD 1 1 1 1 40 1 1 1 1 50 1 1 1 1 1  $\mathbf 1$ 17 60 1 1 1 1 50 1 1 1 45 1 1 1 1 50 1 1 1 1 60 Tmin 1 1 1  $\mathbf 1$ <sup>16</sup> <sup>100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>60</sup> Tmin <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>60</sup> Tmin <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>60</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>150</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\mathbf{1}$ 15 150 1 1 1 1 1 90 1 1 1 80 1 1 1 1 80 1 1 1 1 200 02,Tmax 1 1 1  $\mathbf{1}$ <sup>14</sup> <sup>200</sup> O2 <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>140</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>100</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> <sup>250</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\overline{1}$ 13 250 1 1 1 250 1 1 1 1 150 1 1 1 1 1 1 1 1 1 1 1 1 300 1 1 1 1  $\overline{1}$ 12 300 Tmax 1 1 1 1 300 Tmax 1 1 1 200 1 1 1 1 200 1 1 1 1 1 400 1 1 1 1  $\mathbf{1}$ 11 500 Salmax 1 1 1 1<br>10 750 1 1 1 1 1 1 1 1 250 1 1 1 250 1 1 1 1 500 1 1 1 1 1 10 750 1 1 1 1 1 1 1 1 300 1 1 1 1 300 1 1 1 1 750 1 1 1 1 9 1000 1 1 1 1  $1000$  1000 1 1 1 1 8 1250 1 1 1 1 <sup>1250</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> 7 1500 1 1 1 1 <sup>1500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> 6 2000 1 1 1 1  $\frac{1}{2000}$  1 1 1 1 5 2500 1 1 1 1  $\frac{1}{2500}$  1 1 1 1 4 3000 1 1 1 1 **1** 3000 1 1 1 1 <sup>3</sup> <sup>3500</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> $\frac{1}{3500}$  1 1 1 1 2 3930 1 1 1 1 <sup>4030</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup> CTD006 wp0210 CTD007 wp0211 CTD008 wp02122 CTD010 wp0301 CTD010 wp0302 CTD010 wp0302 CTD001 wp0205 CTD002 wp0206 CTD003 wp0207CTD003 wp0207 CTD004 wp0208 CTD005 wp0209 CTD005 wp0209

**Table 3: Summary of hydrochemistry samples collected (salinity, dissolved oxygen, nutrients) from V3 KAXIS CTDs 1-47. Note that nutrients were collected in triplicate.** 

<sup>4080</sup> <sup>1</sup> <sup>1</sup> <sup>1</sup>

1 3970 1 1 1 1







